

A-370 Multiplexed, Isotype-Specific Ultrasensitive Research-Use Bridging Serology Assays for Detection of Autoimmune Reactivities

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1 Abstract

Background: Autoimmune diseases affect over 50 million Americans and have rapidly rising incidence. The presence of multiple specific autoantibodies can often predict disease onset in at-risk individuals [e.g. type 1 diabetes (T1D), systemic lupus erythematosus, celiac disease], and assist in distinguishing disorders with similar clinical features (e.g. T1D versus type 2 diabetes). In an ongoing study to assess the feasibility of developing highly sensitive and specific multiplexed antigen-bridging serology panels for detection of autoimmune biomarkers, the MSD® MULTI-ARRAY technology is being used to produce research-use assays for detection of T1D, other organ-specific autoimmune disease, and connective tissue disorder biomarkers. First generation multiplexed assay panels developed to detect T1D autoantibodies to glutamic acid decarboxylase (GADA), insulinoma 2 (IA2A), and insulin (IAA), as well as markers relevant to celiac disease, autoimmune gastritis, and thyroid disease, were previously tested using assay proficiency evaluation samples from the Islet Autoantibody Standardization Program. In those studies, the MSD T1D-relevant assays performed comparably to existing assays, with the advantages of low sample requirements for multiplexed detection (<25 µL needed for detection of up to 10 biomarkers), high throughput, and no radioactivity [Mathew et al. (2016) Diabetes 65(Supplement 1):A431-A440, 1673P]. In the current phase of the project, next generation assays have been developed using MSD's ultrasensitive assay format to enhance multiplexed detection of autoimmune disease-related reactivities with the additional capability to discern antibody isotypes.

Objective: The objective of the study was to compare performance of the first and second generation MSD bridging serology assay panels in a 96-well plate, high throughput format. The assays were assessed using commercially purchased T1D and presumably normal individual sera. The multiplexing capability enabled simultaneous quantitative measurement of T1D and comorbid disease markers. The second generation assay was formatted for detection of IgG reactivities, but can easily be formatted for detection of other antibody isotypes (IgA, IgM).

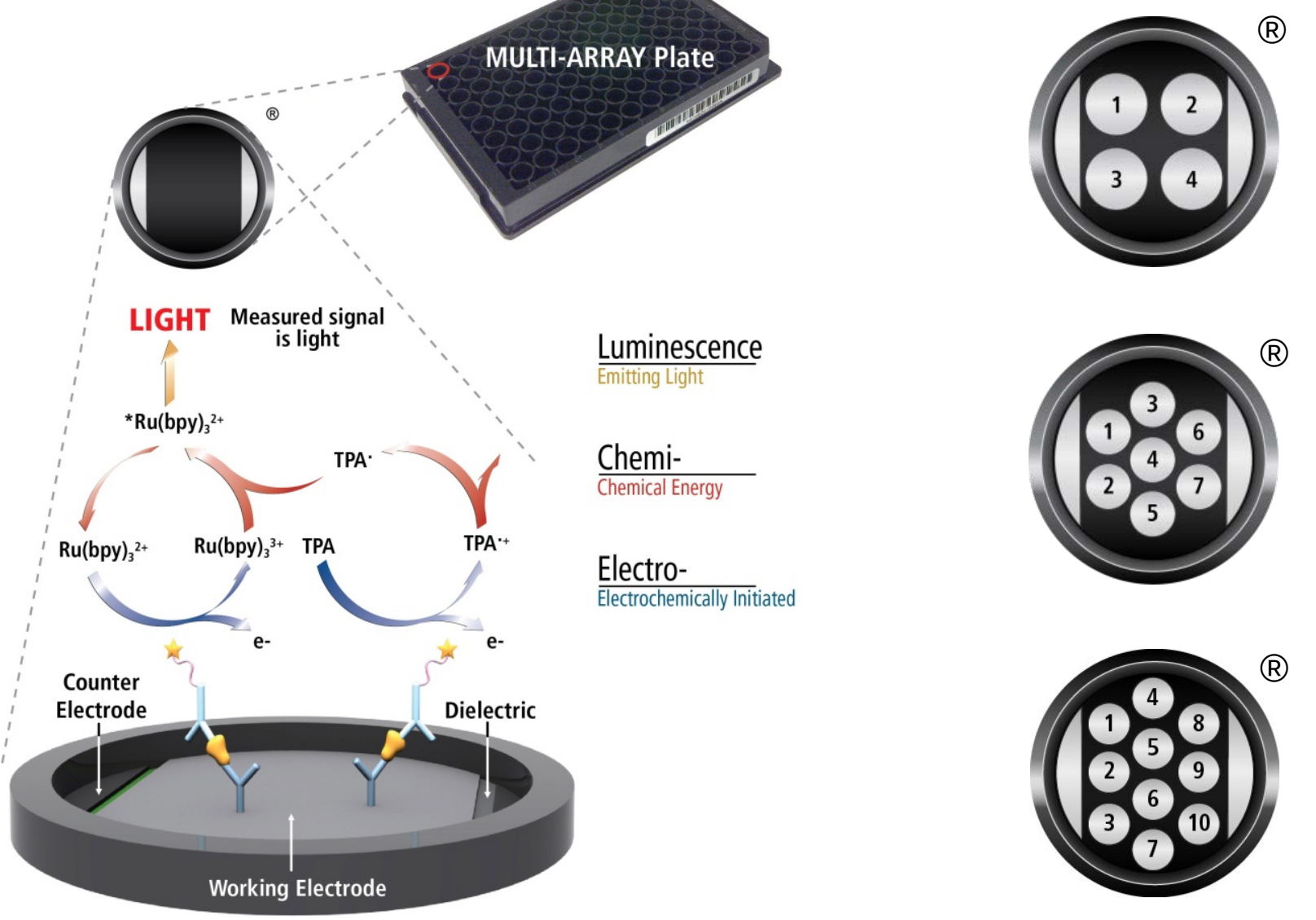
Results: Preliminary data show that assay sensitivities/specificities improve from 65%/100% to 91%/96% (IAA detection), 57%/100% to 70%/92% (GADA detection), and 35%/95% to 48%/96% (IA2A detection) when comparing the first and second generation assays, respectively. Furthermore, ultrasensitive assays were included for detection of pro-insulin autoantibodies (pro-IAA) and zinc transporter 8 (ZnT8) that demonstrated initial specificity/sensitivity values of 96%/100% and 22%/92%, respectively. Two or more T1D reactivities were detected in eighteen of the twenty-four T1D samples tested and in one of the twenty-five normal samples screened.

Conclusion: The sample-sparing ultrasensitive multiplexed MSD autoantibody assays significantly enhance the ability to identify samples containing multiple autoimmune reactivities, with the capability to distinguish specific antibody isotypes, while retaining the high specificity of the bridging serology assay format.

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2 MSD Technology

MSD's electrochemiluminescence detection technology uses SULFO-TAG™ labels that emit light upon electrochemical stimulation initiated at the electrode surfaces of MULTI-ARRAY® and MULTI-SPOT® microplates.



3 Methodology and Preliminary Data

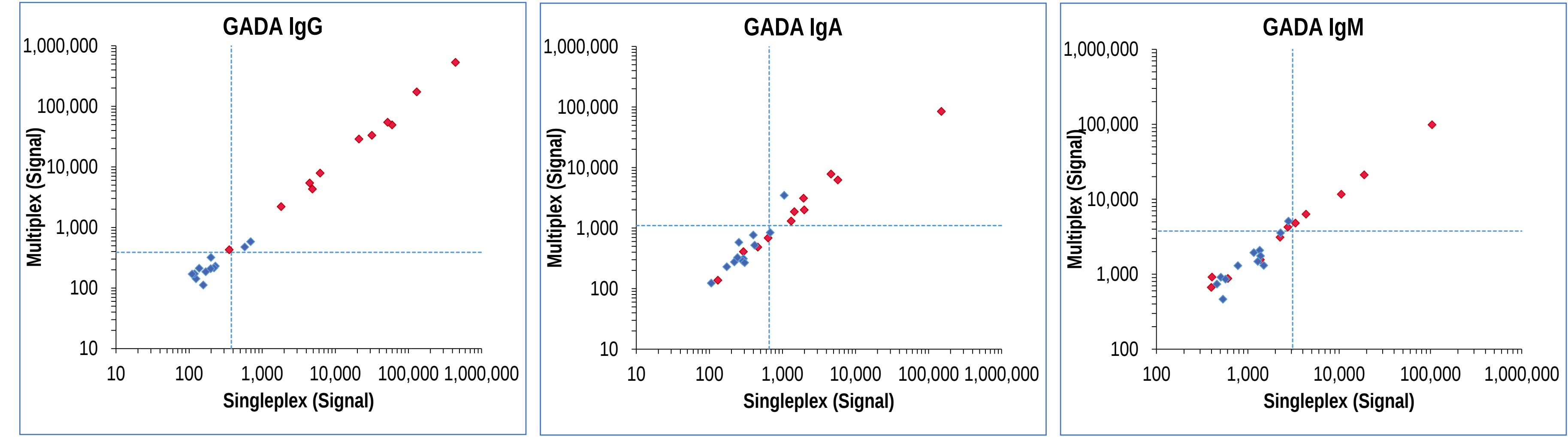
The bridging assay format is used to detect antigen-specific autoantibodies in serum samples. This method uses autoantigens labeled with either detection or capture tags. Antigen-specific antibodies must bind to both species of antigen to obtain signal, ensuring improved assay specificity. Using U-PLEX® technology, we combine the performance advantages of the bridging format with the sample-sparing advantages of MSD multiplexed assays. Simultaneous detection of multiple autoantigen reactivities minimizes the amount of sample needed. Samples are tested in duplicate, along with MSD positive control sera, where available. First generation multiplexed bridging assays for T1D-relevant biomarkers have previously been demonstrated to show excellent discrimination of positive from negative reactive samples using the IASP blinded sample cohort. The current generation of assays incorporates an optimized assay format and adds the ability to distinguish antibody isotypes. Preliminary data were generated using commercially purchased T1D and normal serum samples, prior to full assay optimization, generating the data provided in the abstract and below.

	IAA		GADA		IA2A		pro-IAA	ZnT8A
	1 st Gen	2 nd Gen (IgG)	1 st Gen	2 nd Gen (IgG)	1 st Gen	2 nd Gen (IgG)	2 nd Gen (IgG)	2 nd Gen (IgG)
Sensitivity	65%	91%	57%	70%	35%	48%	96%	22%
Specificity	100%	96%	100%	92%	95%	96%	100%	92%

Isotype-specific MSD bridging serology assays can be multiplexed with minimal effects on assay performance. Data are shown for the IgG, IgA, and IgM GADA reactivities tested in singleplex, or multiplexed with IAA, pro-IAA, and ZnT8A, using commercially purchased normal and T1D serum samples.

Dashed lines in the figures below denote assay cutoffs determined as median signal of normal samples + 2.2 x interquartile range (IQR).

Isotype-Specific Bridging Immunoassay for GADA – Multiplex Versus Singleplex Assay Formats



4 Sample Testing Results

Second generation isotype-specific bridging serology assays to detect the T1D-relevant IAA, pro-IAA, ZnT8A, GADA, and IA2A reactivities were further optimized and combined with newly developed assays to measure autoantibodies to Transglutaminase (TGA, celiac disease), Deamidated Gliadin Peptides (DGPA, celiac disease), Thyroglobulin (TGBA, autoimmune thyroid disease), 17-Hydroxylase (17-OHA, Addison's disease), Jo-1 (Jo-1A, polymyositis), and Intrinsic Factor (IFA, pernicious anemia). These assays were multiplexed in panels based on assay compatibilities and required sample dilutions. Samples were tested for IgG and IgA reactivities in duplicate, using less than 50 µL of each serum sample for all measurements.

- The serum samples from the University of Florida used for testing were:
- 96 T1D or other autoimmune disease biomarker positive samples characterized for individual reactivities (levels of other reactivities and disease status not defined).
 - 16 positive samples each for IAA (University of Florida in-house macro-IAA RIA), GADA (Kronus RIA), and IA2A (Kronus RIA)
 - 18 ZnT8A positive samples (Kronus ELISA)
 - 10 positive samples each for TGA (Kronus RIA), Thyroid Peroxidase (TPOA, Kronus RIA), and adrenal cell autoantibodies (ACA, LDT indirect immunofluorescence using human adrenal tissue)
- (2) 64 age/gender matched normal serum samples. These samples have not been screened for T1D or other autoimmune disease markers and are hence uncharacterized.

Average sample signal data are summarized at right, with signal intensities observed spanning 3-4 orders of magnitude. Sample signals above each assay's cutoff signal value (median + 2.2 x IQR) are highlighted in red font.

The results demonstrate that 85% of the individual biomarker positive samples exhibit more than one immune reactivity, with varied combinations of markers observed. Of the provided T1D marker positive samples, 97% show multiple autoimmune reactivities, and 92% are positive for more than one T1D-specific marker. The magnitudes of the signals observed in the marker positive samples are much higher than in the normal serum samples.

Among the uncharacterized normal samples, 34% exhibit multiple reactivities, and 14% have multiple T1D-specific markers.

The percent of samples positive for each marker (IgG, IgA, or combined isotypes), in each sample subset are summarized below. The percent of samples with multiple T1D markers is shown for each sample subgroup.

		Samples (Percent Above Cutoff)									
Assay	Isotype	IAA positive (n=16)	ZnT8A positive (n=18)	IA2A positive (n=16)	GADA positive (n=16)	TGA positive (n=10)	TPOA positive (n=10)	ACA positive (n=10)	Normal (n=64)		
IAA	IgG	88	50	44	69	0	10	40	13		
	IgA	25	11	0	31	10	20	40	13		
pro-IAA	IgG + IgA	94	61	44	75	10	30	50	17		
	IgG	69	39	38	44	0	0	50	2		
ZnT8A	IgG + IgA	19	22	19	44	10	30	20	9		
	IgG	69	50	50	63	10	30	70	11		
IA2A	IgG + IgA	56	83	44	31	0	0	10	5		
	IgG	63	72	88	56	0	10	30	2		
GADA	IgG + IgA	50	67	69	50	0	20	10	17		
	IgG	69	72	88	56	0	30	30	19		
TGA	IgG	0	11	6	13	60	40	40	5		
	IgG + IgA	25	22	6	6	40	30	20	5		
DGPA	IgG	25	22	6	13	60	40	40	8		
	IgG + IgA	31	27	13	19	0	10	20	16		
TGBA	IgG	19	28	25	31	0	20	10	14		
	IgG + IgA	31	39	31	38	0	20	20	23		
17-OHA	IgG	19	0	13	38	40	30	30	3		
	IgG + IgA	19	6	13	19	10	50	0	8		
Jo-1A	IgG	19	17	6	38	40	10	0	6		
	IgG + IgA	19	11	6	13	0	10	10	8		
IFA	IgG	25	22	13	44	40	10	10	14		
	IgG + IgA	19	39	38	19	10	20	30	11		
% Samples with >1 T1D Reactivity	IgG	13	22	19	19	0	0	0	2		
	IgG + IgA	13	17	6	13	20	30	0	3		
	IgG	25	28	25	25	20	30	0	3		
	IgG + IgA	25	28	25	25	20	30	0	3		

	IAA	pro-IAA		ZnT8A		IA2A		GADA		TGA		DGPA		TGBA		17-OHA		Jo1A		IFA			
		Signal	IQR	Signal	IQR	Signal	IQR	Signal	IQR	Signal	IQR	Signal	IQR	Signal	IQR	Signal	IQR	Signal	IQR	Signal	IQR		
IAA Positive Samples	1,382	380	159	250	94,826	11,561	127,446	15,991	112,213	14,147	147	141	85,748	8,669	4,159	62,827	4,429	12,213	1,832	385	79	127	
	79,297	3,975	4,632	966	4,490	4,462	21,496	1,477	1,386,844	11,204	6	151	439	179	145	149	8,660	967	401	482	179	145	
	1,014	2,693	293	356	2,440	3,538	96,979	1,659	1,879	402	32	6	368	735	85	75	2,442	4,794	612	633	122	1,937	
	1,481	377	19	122	8,727	1,603	239,624	1,905	1,320	42,821	198	106	106	8,242	1,062	646	3,407	2,958	499	688	1,238	275	
	2,389	291	727	109	1,616,599	127,446	1,739,288	149,888	1,463,679	18,991	77	118	87,838	18,442	548	161,074	2,952	1,561	1,947	637	235	140	
	1,479	372	419	118	1,606	1,979	2,140	1,477	1,606	4,639	198	106	64	951	60	302	654	257	364	127	132		
	755	756	238	100	1,406	724	49,540	31	2,097	184	46	384	99	102	39	42	859	254	402	371	173	70	
	284	278	331	98	1,481	953	498	323	262	114	60	127	29	155	88	119	399	360	307	315	93	65	
	931	901	147	98	2,379	1,853	600	358	280	147	115	112	1,212	1,200	61	556	760	371	384	165	127		
	8,266	1,152	107	3,120	1,431	8,637	1,654	99	1,654	99	109	109	1,173	470	109	1,173	470	109	1,173	470	109	1,173	
ZnT8A Positive Samples	269	3,381	146	272	1,593	2,651	687	449	169	605	65	1,382	137	422	47	152	244	1,219	371	815	68	290	
	308	499	231	269	1,229	2,209	344	297	159	265	144	144	209	265	265	265	265	265	265	265	265	265	
	687	7,005	258	288	2,822	4,860	1,712,184	1,883,337	706	269	706	269	44	157	188	135	206	1,084	2,897	1,081	96	144	
	133	202	94	110	11,847	1,857	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	
	214	402	105	110	16,684	2,795	3,179	1,288	13,767	1,614	51	63	230	109	103	5,992	1,406	4,021	1,020	97	144		
	755	756	238	100	1,406	724	49,540	31	2,097	184	46	384	99	102	39	42	859	254	402	371	173	70	
	284	278	331	98	1,481	953	498	323	262	114	60	127	29	155	88	119	399	360	307	315	93	65	
	931	901	147	98	2,379	1,853	600	358	280	147	115	112	1,212	1,200	61	556	760	371	384	165	127		
	8,266	1,152	107	3,120	1,431	8,637	1,654	99	1,654	99	109	109	1,173	470	109	1,173	470	109	1,173	470	109	1,173	
	269	3,381	146	272	1,593	2,651	687	449	169	605	65	1,382	137	422	47	152	244	1,219	371	815	68	290	
IA2A Positive Samples	308	499	231	269	1,229	2,209	344	297	159	265	144	144	209	265	265	265	265	265	265	265	265	265	
	687	7,005	258	288	2,822	4,860	1,712,184	1,883,337	706	269	706	269	44	157	188	135	206	1,084	2,897	1,081	96	144	
	133	202	94	110	11,847	1,857	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	
	214	402	105	110	16,684	2,795	3,179	1,288	13,767	1,614	51	63	230	109	103	5,992	1,406	4,021	1,020	97	144		
	755	756	238	100	1,406	724	49,540	31	2,097	184	46	384	99	102	39	42	859	254	402	371	173	70	
	284	278	331	98	1,481	953	498	323	262	114	60	127	29	155	88	119	399	360	307	315	93	65	
	931	901	147	98	2,379	1,853	600	358	280	147	115	112	1,212	1,200	61	556	760	371	384	165	127		
	8,266	1,152	107	3,120	1,431	8,637	1,654	99	1,654	99	109	109	1,173	470	109	1,173	470	109	1,173	470	109	1,173	
	269	3,381	146	272	1,593	2,651	687	449	169	605	65	1,382	137	422	47	152	244	1,219	371	815	68	290	
	308	499	231	269	1,229	2,209	344	297	159	265	144	144	209	265	265	265	265	265	265	265	265	265	265
TGA Positive Samples	687	7,005	258	288	2,822	4,860	1,712,184	1,883,337	706	269	706	269	44	157	188	135	206	1,084	2,897	1,081	96	144	
	133	202	94	110	11,847	1,857	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641
	214	402	105	110	16,684	2,795	3,179	1,288	13,767	1,614	51	63	230	109	103	5,992	1,406	4,021	1,020	97	144		
	755	756	238	100	1,406	724	49,540	31	2,097	184	46	384	99	102	39	42	859	254	402	371	173	70	
	284	278	331	98	1,481	953	498	323	262	114	60	127	29	155	88	119	399	360	307	315	93	65	
	931	901	147	98	2,379	1,853	600	358	280	147	115	112	1,212	1,200	61	556	760	371	384	165	127		
	8,266	1,152	107	3,120	1,431	8,637	1,654	99	1,654	99	109	109	1,173	470	109	1,173	470	109	1,173	470	109	1,173	
	269	3,381	146	272	1,593	2,651	687	449	169	605	65	1,382	137	422	47	152	244	1,219	371	815	68	290	
	308	499	231	269	1,229	2,209	344	297	159	265	144	144	209	265	265	265	265	265	265	265	265	265	265
	687	7,005	258	288	2,822	4,860	1,712,184	1,883,337	706	269	706	269	44	157	188	135	206	1,084	2,897	1,081	96	144	
TPOA Positive Samples	133	202	94	110	11,847	1,857	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641
	214	402	105	110	16,684	2,795	3,179	1,288	13,767	1,614	51	63	230	109	103	5,992	1,406	4,021	1,020	97	144		
	755	756	238	100	1,406	724	49,540	31	2,097	184	46	384	99	102	39	42	859	254	402	371	173	70	
	284	278	331	98	1,481	953	498	323	262	114	60	127	29	155	88	119	399	360	307	315	93	65	
	931	901	147	98	2,379	1,853	600	358	280	147	115	112	1,212	1,200	61	556	760	371	384	165	127		
	8,266	1,152	107	3,120	1,431	8,637	1,654	99	1,654	99	109	109	1,173	470	109	1,173	470	109	1,173	470	109	1,173	
	269	3,381	146	272	1,593	2,651	687	449	169	605	65	1,382	137	422	47	152	244	1,219	371	815	68	290	
	308	499	231	269	1,229	2,209	344	297	159	265	144	144	209	265	265	265	265	265	265	265	265	265	265
	687	7,005	258	288	2,822	4,860	1,712,184	1,883,337	706	269	706	269	44	157	188	135	206	1,084	2,897	1,081	96	144	
	133	202	94	110	11,847	1,857	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641
ACA Positive Samples	214	402	105	110	16,684	2,795	3,179	1,288	13,767	1,614	51	63	230	109	103	5,992	1,406	4,021	1,020	97	144		
	755	756	238	100	1,406	724	49,540	31	2,097	184	46	384	99	102	39	42	859	254	402	371	173	70	
	284	278	331	98	1,481	953	498	323	262	114	60	127	29	155	88	119	399	360	307	315	93	65	
	931	901	147	98	2,379	1,853	600	358	280	147	115	112	1,212	1,200	61	556	760	371	384	165	127		
	8,266	1,152	107	3,120	1,431	8,637	1,654	99	1,654	99	109	109	1,173	470	109	1,173	470	109	1,173	470	109	1,173	
	269	3,381	146	272	1,593	2,651	687	449	169	605	65	1,382	137	422	47	152	244	1,219	371	815	68	290	
	308	499	231	269	1,229	2,209	344	297	159	265	144	144	209	265	265	265	265	265	265	265	265	265	265
	687	7,005	258	288	2,822	4,860	1,712,184	1,883,337	706	269	706	269	44	157	188	135	206	1,084	2,897	1,081	96	144	
	133	202	94	110	11,847	1,857	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641	1,641
	214	402	105	110	16,684	2,795	3,179	1,288	13,767	1,614	51	63	230	109	103	5,992	1,406	4,021	1,020	97	144		
Unlabeled Normal Serum Samples	755	756	238	100	1,406	724	49,540	31	2,097	184	46	384	99	102	39	42	859	254	402	371	173	70	
	284	278	331	98	1,481	953	498	323	262	114	60	127	29	155	88	119	399	360	307	315	93	65	