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Abstract

Apoptosis, or programmed cell death, plays a critical role in the control of disease and is the target of current drug discovery efforts to improve the efficacy of cancer therapeutics. Pro-survival pathways are similarly important in maintaining normal cell function where defects also lead to disease. Apoptosis and cell survival are both regulated through a variety of pathways acting in parallel that utilize phosphorylation as a key regulatory mechanism. However, limited approaches are available for simultaneously measuring key phosphorylated intermediaries in both of these pathways in a high throughput manner. Here we demonstrate the ability to detect simultaneously panels of the phosphorylated proteins Akt, GSK-3 cx, Bad, and p53 in a sensitive, quantifiable assay format. Additionally, the levels of phosphorylated protein can be compared to their total level; we demonstrate this measurement with the quantitation of phosphorylated and total Akt in Jurkat whole cell lysates in the same well. Both assays afford fast, simple protocols in which the results obtained from treated and untreated cells agree with those obtained by traditional western blot analysis. We think these measurements provide new tools towards the comprehensive understanding of signaling pathways in diseased and normal tissue.

Apoptosis Phosphoprotein Assay Format



Protocol

- 1. MSD MULTI-SPOT[™] 4 Spot 96-Well plates precoated with capture antibodies are blocked with 3% BSA in TBS buffer (150mM NaCl, 50mM Tris-HCl pH7.5), 50µL per well, 2h. Wash with TBS
- 2. Cell lysates are incubated in the assay plate for 1h with shaking, 25µL per well. Lysate diluent: TBS buffer with fresh phosphatase inhibitor cocktails 1 and 1I, and a protease inhibitor cocktail. Wash with TBS
- 3. Antibodies labeled with MSD SULFO-TAG[™] in TBS buffer with 1% MSD Blocker A are pre-mixed and incubated in the assay plate for 1h with shaking, 25µL per well. Wash with TBS
- 4. MSD Read Buffer T (IX), ISOµL per well, followed by plate analysis on an MSD SECTOR Imager instrument.



Detection of Phosphorylated Akt in Whole Cell Lysates





Logarithmically growing Jurkat cells were treated with Ly inhibitor for 1h. Whole cell lysates were added to MSD MULTI-SPOT 4 Spot 96-well plates coated with anti-pan-Akt antibody on one of the four spatially distinct electrodes per well. BSA was coated onto the remaining three electrodes in each well. Phosphorylated Akt was detected with 10nM anti-phospho-Akt antibody labeled with MSD SULFO-TAG reagent.

Lysate (µg)	Akt lysat	es (untrea	ted cells)	Akt lysates (treated cells)			S-B	S/B
	Ave ECL	Std.Dev.	%CV	Ave ECL	Std.Dev.	%CV		
0	45	5	11	37	7	19	8	1.2
0.5	200	23	12	39	I	4	161	5.1
I	393	6	Ι	64	6	9	329	6.1
5	2,738	134	5	127	4	3	2,611	21.6
10	8,196	145	2	169		6	8,027	48.6
15	14,485	406	3	261	25	10	14,224	55.5
20	19,034	7	0	327	6	2	18,708	58.3



Detection of Phosphorylated Bad in Whole Cell Lysates



20

45,531

496

9,202

18

0

36,329

4.9



Logarithmically growing COS-7 cells were serum-starved overnight, followed by treatment with PMA for 1h. Whole cell lysates were added to MSD MULTI-SPOT 4 Spot 96-well plates coated with antiphospho-Bad antibody on one of the four spatially distinct electrodes per well. BSA was coated onto the remaining three electrodes in each well. Phosphorylated Bad was detected with 10nM anti-total-Bad antibody labeled with MSD SULFO-TAG reagent.



\odot Detection of Phosphorylated GSK-3lpha in Whole Cell Lysates





Logarithmically growing Jurkat cells were treated with staurosporine for 4h. Whole cell lysates were added to MSD MULTI-SPOT 4 Spot 96-well plates coated with anti-phospho-GSK-3 α antibody on one of the four spatially distinct electrodes per well. BSA was coated onto the remaining three electrodes in each well. Phosphorylated GSK-3 α was detected with 10nM anti-total-GSK-3 α antibody labeled with MSD SULFO-TAG reagent.



• Multiplex p53 Assay: Detection of Phosphorylated and Total p53 in the Same Well





Media was removed from logarithmically growing HT29 cells, followed by UV irradiation at 40mJ/cm². Whole cell lysates were added to MSD MULTI-SPOT 4 Spot 96-well High Bind plates coated with antiphospho-p53 antibody and anti-total-p53 antibody coated on spatially distinct electrodes in the same well. BSA was coated onto the remaining two electrodes in each well. Phosphorylated and total p53 were detected with 5nM anti-total-p53 antibody labeled with MSD SULFO-TAG reagent. A titration of HT29 cell lysates shows detection of increasing phosphorylated p53 while the ratio of treated/untreated total p53 remains constant.

Lysate (µg)	р-р5	3 (treated	cells)	р-р53	S/B		
	Ave	Std.Dev.	%CV	Ave	Std.Dev.	%СV	
0	141	4	3	165	5	3	0.9
0.3125	8,506	132	2	7,316	612	8	1.2
0.625	14,904	474	3	10,418	1,095	- 11	1.4
1.25	20,206	482	2	10,422	1,111	11	1.9
2.5	24,992	2,850	11	8,035	1,323	16	3.1
5	40,285	611	2	8,585	48		4.7
10	53,398	2,809	5	9,011	127	I	5.9
20	81,312	462		11,525	531	5	7.1

Phospho-p53

Total p53									
Lysate (µg)	p53	(treated o	:ells)	p53 (p53 (untreated cells)				
	Ave	Std.Dev.	%CV	Ave	Std.Dev.	%CV	Untreated		
0	695	187	27	588	98	17	1.2		
0.3125	24,204	1,877	8	24,369	4,078	17	1.0		
0.625	46,860	986	2	50,236	10,960	22	0.9		
1.25	76,646	3,594	5	79,023	29,241	37	1.0		
2.5	120,236	8,567	7	129,232	25,356	20	0.9		
5	147,757	18,393	12	197,627	14,181	Ι	0.7		
10	167,174	4,006	2	252,273	9,170	8	0.7		
20	257,884	66,759	26	316,840	60,796	19	0.8		



Multiplex Akt Assay: Detection of Phosphorylated and Total Akt in the Same Well

BSA

Total

Akt



Logarithmically growing Jurkat cells were treated with Ly inhibitor for 1h. Whole cell lysates were added to MSD MULTI-SPOT 4 Spot 96-well plates coated with antiphospho-Akt antibody and anti-total-Akt antibody coated on spatially distinct electrodes in the same well. BSA was coated onto the remaining two electrodes in each well. Phosphorylated and total Akt were detected with 10nM anti-total-Akt antibody labeled with MSD SULFO-TAG reagent. A titration of Jurkat cell lysates shows detection of increasing phosphorylated Akt while untreated/treated for total Akt remains constant.

Phos	pho-	Akt

Lysate (µg)	p-Akt lysates (untreated cells)			p-Akt lysates (treated cells)			S-B	S/B
	Ave	Std.Dev.	%СV	Ave	Std.Dev.	%CV		
0	292	27	9	322	44	14	-30	0.9
5	6,592	445	7	952	11	I	5,640	6.9
10	12,832	49	0	1,356	74	5	11,476	9.5
20	21,964	236	I	2,056	124	6	19,878	10.5

Total-Akt								
Lysate (µg)	Akt lysat	es (untrea	Untreated/Treated					
	Ave	Std.Dev.	%CV	Ave	Std.Dev.	%СV		
0	154	20	13	131	I	I	1.2	
5	17,347	854	5	22,577	3,340	15	0.8	
10	26,362	151	Ι	36,862	956	3	0.7	
20	41,392	2,100	5	54,564	1,625	3	0.8	



Multiplex Apoptosis Panel: Detection of FOUR Phosphoproteins in the Same Well

pSer473

Akt

pSer15

p53

Reporter antibodies:

10nM anti-phospho-Akt-SULFO-TAG antibody,

10nM anti-Bad-SULFO-TAG antibody,

10nM anti-GSK-3 &-SULFO-TAG antibody, and 5nM anti-p53-SULFO-TAG antibody

Whole cell lysates were added separately to MSD MULTI-SPOT 96-Well 4 Spot plates pre-coated with anti-Akt, anti-Gsk-3 α , anti-p53, and anti-Bad antibodies immobilized on four spatially distinct electrodes in a single well. Phosphorylated proteins were detected with reporter antibodies labeled with MSD SULFO-TAG reagent.



Results with Jurkat whole cell lysates show that only phosphorylated Akt and GSK-3 α are detected. Phosphorylated p53 is detected with HT29 cell lysates, but not any of the other three markers. COS-7 cell lysates contain endogenous p53, shown in the western blot and confirmed in this multiplex assay. Therefore, we predict and have demonstrated here that the data and corresponding images show the same level of endogenous p53 for both treated and untreated COS-7 cell lysates, in addition to detection of phosphorylated Bad for treated cells. Individual measurements from each of the four spots remained highly analytical as regards sensitivity and precision and showed no spurious crosstalk between putative measurements, underscoring the utility of the method.



Conclusions

A panel of multiplex apoptotic phosphoprotein assays was developed to detect phospho-Akt, phospho-GSK-3 α , phospho-p53 and phospho-Bad in whole cell lysates using MSD's MULTI-SPOT 4 Spot plates. The system allowed us to evaluate simultaneously several key regulatory proteins in the apoptosis and cell survival pathways. First, we demonstrated that the four phosphoproteins could be individually measured with very high specificity from select cell types. Besides detecting Akt from growing Jurkat cells, the key downstream partners to the Akt signaling pathway acting to prevent programmed cell death, phospho-Bad and phospho-GSK-3 α were detected from PMA treated COS-7 cells and staurosporine treated Jurkat cells, respectively, with high signal to background ratios and excellent precision. As expected, when phospho-Akt was suppressed selectively by the Pl3K specific LY294002 inhibitor, total Akt in these cells did not change as demonstrated by our duplex assay that simultaneously detected the two forms in a single well. Parallel to this, a duplex assay was developed to detect phospho and total forms of p53 from UV treated HT29 cells. The detection assays were further multiplexed to accommodate all four phosphoprotein targets in a single well. Protocols for MSD multiplex assays are fast, simple and boast the sensitivity and specificity observed in traditional western blot analysis with whole cell lysates. The specific detection of each protein in these multiplex panels presents a versatile analytical tool for determining the status of these proteins with high precision. These methods should be useful in elucidating mechanisms of apoptosis and, more generally, in checking and cross checking the status of various signaling pathways in the presence of stimuli.

