Development and Characterization of Th17 Related U-PLEX® Assays

Qian Ning, Arun Shrestha, Chris Shelburne, Pu Liu, Mikhael Wallowitz, David Stewart, Robert Wolfert, Pankaj Oberoi, and Jacob N. Wohlstadtner
Meso Scale Discovery, Rockville Maryland, USA

Abstract
Th17-related cytokines mediate a host’s defense mechanisms against various infections and play a crucial role in crosstalk between the immune system and affected tissues. Here we describe the development and characterization of multiplexed Th17-related immunosassays on MSD’s flexible U-PLEX platform. Monoclonal and polyclonal antibodies were conjugated with biotin and/or SULFO-TAG™ and screened as capture and/or detection reagents. A number of analytical parameters were used to select antibody pairs, and assays were developed by optimizing antibody concentrations, calibrator curve, specificity, matrix tolerance, and assay robustness. Assay performance was verified to be compatible with other U-PLEX assays by running Th17-related multiplexes on the U-PLEX platform with controls and samples.

Calibration curves showed expected signals, sensitivity, precision, and accuracy. Control samples for the assays had CVs < 10% within runs and < 25% between runs. Sensitivities were < 1 pg/mL for the majority of the assays. Dilution linearity and spike recovery studies demonstrated acceptable matrix tolerance and accurate quantification for most of the assays tested across all matrices (typically between 75% - 125%). Cross-reactivity between assays was shown to be typically < 0.5%. Results demonstrated a strong correlation between samples measured on U-PLEX multiplex and streptavidin plates with r² values > 0.95 and slopes between 0.98-1.2

Over 20 Th17-related assays for human and mouse were developed for the U-PLEX platform. These assays can make biological measurements on matrices that are relevant to a wide range of life science applications and are used as part of our pre-clinical or clinical studies.

Methods
MSD’s electrochemiluminescence (ECL) detection technology uses SULFO-TAG labels that emit light upon electrochemical excitation initiated at the electrode surfaces of MULTI-ARRAY® and MULTI-SPOT® microplates. The U-PLEX assay platform utilizes ten unique linkers that specifically bind to individual spots enabling simple and flexible creation of multiplex immunosassays.

Calibration Curves and Limits of Detection
The figures at right demonstrate typical calibration curves for 12 Th17 related (human) assays. Up to ten markers can be combined per plate to create the desired multiplex.
The table below demonstrates typical data for the 12 assays tested. Data include Hill slopes and lower limits of detection (LLOD) for the calibration curves. The calibrator is reconstituted and diluted serially (4-fold) to generate a 7-point standard curve.

<table>
<thead>
<tr>
<th>Assay</th>
<th>LLOD (pg/mL)</th>
<th>Hill Slope</th>
<th>LLOD (pg/mL)</th>
<th>Hill Slope</th>
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</thead>
<tbody>
<tr>
<td>IL-17A</td>
<td>2.99</td>
<td>1.22</td>
<td>1.23</td>
<td>1.02</td>
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<tr>
<td>IL-17F</td>
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<td>1.45</td>
<td>0.95</td>
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<td>IL-27</td>
<td>1.00</td>
<td>1.56</td>
<td>1.05</td>
<td>1.66</td>
</tr>
</tbody>
</table>

Spike Recovery
Normal human serum (n=5) and EDTA plasma (n=5) samples from a commercial source and cell culture media were spiked with calibrators at three levels (high, mid, and low). % Recovery = (measured concentration – sample endogenous level) / spiked concentration x 100

The percent recovery for most of the assays was within the targeted range (70-130%). IL-33 low recovery was due to high levels of receptor ST2 present in human serum and plasma. IL-29 recovery was found to be sample-dependent with low recovery measured on a different set of samples (data not shown). Additional sample dilution may be used to reduce any matrix effect.

Dilution Linearity
To assess linearity, normal human serum (n=5) and EDTA plasma samples (n=5) from a commercial source, and cell culture media (n=5) were spiked with recombinant calibrator and diluted 1, 2, 4, and 8 fold before testing. The average percent recovery is based on samples that measured within the quantitative range of the assay.

% Recovery = (measured concentration / expected concentration) x 100

The percent recovery for serum samples is shown at right. Both serum and plasma samples were found to be within the targeted range (70-130%) for most of assays, suggesting that the samples dilute linearly from 1-8 fold. IL-33 and IL-17E were non-linear for both serum and plasma matrices, which may be associated with known receptor interferences and inter-sample variation.

Native Sample Testing
Normal human serum (n=8) and EDTA plasma (n=8) samples from a commercial source were tested neat. To demonstrate the detection of native analytes in matrix, cell culture samples (n=8) that were stimulated with different compounds in vitro were also tested. Results for each sample set are displayed at right.

% Recovery = (measured concentration / expected concentration) x 100

All the native analytes are detectable in the stimulated samples; many analytes are detectable in the normal serum and EDTA plasma as well.

Conclusion
The U-PLEX platform enables the rapid design and building of multiplex biomarker panels on the sensitive MSD® platform in a few easy steps. Optimization and characterization data from 12 Th17 related human biomarkers demonstrated sensitivity, precision and accuracy within and between assays.

More broadly, 114 biomarker assays (human: 51; mouse: 27; NHP: 36) have been successfully developed on the U-PLEX platform. Investigators can now combine novel biomarker assays using their own antibodies together with reagents and assays included in the U-PLEX menu. The antibody sets and calibrators provided by MSD have been optimized for performance and ease of use on the U-PLEX platform.