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Quantification of Higher Order Forms of Amyloid Beta and Total Abeta Peptides in CSF

Pankaj Oberoi, Robert Umek, Nyssa Puskar, Jill Dunty, Leonid Dzantiev, David Stewart, and Jacob N. Wohlstadter; Meso Scale Discovery (MSD), Rockville, Maryland, USA **1** Oligomeric Aβ Assay: Interference Study **Φ** Total Aβ Assay **Φ** Peptide-Specific Aβ Assay: Aβ40-Specific Capture

1 Abstract

Background: The Aβ hypothesis of Alzheimer's disease (AD) posits that oligomers and/or aggregates, but not monomers or plaques, are the toxic species that result in neuronal loss. Therefore, assays are needed that can monitor higher order forms in patient samples. Assays that quantify total A β are also in demand for monitoring response to the respective treatment. We report the feasibility of a sandwich immunoassay that binds higher order forms of amyloid beta but not monomers. We also developed an assay that measures total AB. Both assays have been used to characterize CSF samples from healthy controls and patients with various neurological diseases.

Methods: We tested a variety of assay formats with N-terminal, C-terminal, and mid-epitope antibodies against Aß peptides. Monomeric and oligomeric forms of AB40 and AB42 peptides were generated and used to develop the assays. The assay that was designed to measure oligomeric forms used a common N-terminal AB antibody for both capture and detection; the total AB assay used that same antibody for capture but used a mid-epitope antibody for detection. Standard curves presented here show the functionality and specificity of each assay. AB40, AB42, total AB, and oligomeric forms of AB peptide were measured in human CSF.

Results: The total A_β assay had a sensitivity of <5 pg/mL of oligomerized A_{β42}. The oligomeric assay did not detect monomeric A_{β40} but had a sensitivity below approximately 10 pg/mL of oligomerized AB42. (The precise concentration of the oligomerized peptide was difficult to measure.) Although oligomeric forms of AB were not detectable in the CSF samples, roughly 5–20 ng/mL of total AB was detected in the same samples. The total AB levels were greater than the sum of the individual AB40 and AB42 peptides measured independently.

Conclusion: We have developed assays for total and oligomeric forms of AB peptides. The total AB assay was extremely sensitive and could measure differences within a common set of human CSF samples. We were unable to measure oligomeric forms of AB in normal CSF. The development of these assays should enable the evaluation of relative levels AB peptide forms in normal and AD cohorts.

2 Methods



- > MSD's electrochemiluminescence detection technology uses SULFO-TAG[™] labels, which emit light upon electrochemical stimulation initiated at the electrode surfaces of MULTI-ARRAY[®] and MULTI-SPOT[®] microplates
- Higher order forms of AB42 were generated and several assay formats with N-terminal, C-terminal, and mid-epitope antibodies were examined for their ability to measure oligometrized A β 42.
- > All assays used the protocol described below.
- > Human CSF pools (remnant samples) and well-curated human CSF from individual donors were tested with selected assays.
- > Standard curves were analyzed using a 4-parameter logistic model with $1/y^2$ weighting. Error bars = 1 standard deviation (SD).

Protocol

- Add 150 µL of MSD[®] Diluent 35. Incubate for 1 hour at room temperature (RT) with shaking.
- Wash 3X with PBS-T. Add 50 µL of standard or diluted sample. Incubate for 1 hour at RT with shaking.
- Wash 3X with PBS-T. Add 25 µL of detection antibody. Incubate for 1 hour at RT with shaking.
- Wash 3X with PBS-T. Add 150 µL of MSD Read Buffer T. Read on MSD instrument.

3 Higher Order Forms of Aβ Peptides

Agitation and heat were applied to AB40 and AB42 peptides to induce higher order forms. AB peptides were serially diluted (0.375, 0.125, 0.042, and 0.014 mg/mL) and characterized by gel electrophoresis. The SDS-Page gel (Figure A) shows aggregated peptide was formed for AB42, but not for AB40. In contrast, Figure B indicates that AB42 peptides prepared according to MSD manufacturing conditions do not form soluble oligomers but instead display bands at the monomer/tetramer molecular weight marker. The oligomeric AB42 species was used in subsequent assays and the concentration was calculated as 378 ug/mL by a BCA Protein Assay. No other method was available to accurately measure the number of aggregated particles.



The total Aβ assay uses an N-terminal capture antibody (6E10^{*}) and a mid-epitope detection antibody (4G8^{*}) (Figure C). The assay detects and measures both monomeric A β and oligomeric A β species (Figure D). The Hill slopes and sensitivities for oligomeric A β 42, monomeric AB42, and monomeric AB40 are: 1.1 and <5 pg/mL; 1.5 and 1.53 pg/mL; and 1.5 and 1.57 pg/mL, respectively.



6 Oligomeric Aβ Assay

The oligomeric Aβ assay format uses the same N-terminal antibody (6E10) as both the capture and detection antibody (Figure E). This assay format can detect soluble AB oligomers that expose multiple AB N-termini within the higher order structural conformation of aggregated AB. As shown in Figure F, the oligometric assay detects <10 pg/mL of oligometric Aβ42 with a Hill slope of 1.0. The monometric Aβ species did not detect monomeric AB40, but did appear to detect trace amounts of monomeric AB42 with a sensitivity of >100 pg/mL.



6 Peptide-Specific Aβ Assay: Aβ42-Specific Capture

The AB42-specific assay format uses a capture antibody (12F4*) that is reactive to the C-terminus of AB42. The sandwich immunoassay is completed using a detection antibody that is reactive to either the N-terminus (6E10) or mid-region (4G8) of Aβ peptides (Figure G). Both assay formats measured oligomeric AB42 and monomeric AB42, but detected negligible levels of monomeric AB40 (Figures H and I). For the Aβ42-specific assay that used 6E10 detection, Hill slopes and sensitivities are: 1.3 and <10 pg/mL (oligomeric) and 1.4 and 0.67 pg/mL (monomeric). For the AB42-specific assay using 4G8 detection, Hill slopes and sensitivities are: 1.5 and <100 pg/mL (oligomeric) and 1.7 and 1.77 pg/mL (monomeric).











The AB40-specific assay format uses a capture antibody that is reactive to the C-terminus of AB40. The sandwich immunoassay is completed using a detection antibody that is reactive to either the N-terminus (6E10) or mid-region (4G8) of AB peptides (Figure J). Both assay formats measured monomeric AB40, but did not detect monomeric AB42 (Figures K and L). AB42 oligomers are less than 0.01% detectable compared to monomeric AB40. In the graphs below, the red star designates the upper limit of physiological levels of AB40 in human CSF. For the Aβ40-specific assay, the Hill slopes and sensitivities for monomeric Aβ40 are: 1.6 and 1.96 pg/mL (6E10 detection) and 1.5 and 2.52 pg/mL (4G8 detection).



8 Peptide-Specific Aβ Assay: Aβ38-Specific Capture

The AB38-specific assay format uses a capture antibody that is reactive to the C-terminus of AB38. The sandwich immunoassay is completed using a detection antibody that is reactive to either the N-terminus (6E10) or mid-region (4G8) of AB peptides (Figure M). Neither assay detected monomeric AB42 or monomeric AB40 (Figures N and O). AB42 oligomers are not detected within physiologically relevant concentrations; the red star designates the upper limit of physiological levels of AB38 in human CSF. We speculate that high concentrations of aggregated A β 42 bind non-specifically in both the A β 38- and A β 40-specific capture assays.

9 Total Aβ Assay: Aβ Peptide Titration Study

An A β peptide titration study was used to assess the ability of the total A β assay to detect various combinations of monomeric A β 42 and monomeric AB40 peptides. The measured concentration (i.e., apparent AB40, ng/mL) was determined for: (1) a constant level of AB42 (0.3) ng/mL) spiked with various concentrations of AB40 (0, 0.5, 2.5, and 10 ng/mL) and (2) a constant level of AB40 (0.3 ng/mL) spiked with various concentrations of AB42 (0, 0.5, 2.5, and 10 ng/mL). In both cases, total AB, as determined by the sum of monomeric AB42 and monomeric AB40, increased in an additive manner, as measured with the total AB assay. NOTE: The concentrations designated with an asterisk in the table below represent measured concentrations that are above the limit of quantitation.

| Spike Conc. (ng/mL) | | Massured Cons. (ng/ml) |
|----------------------|----------------------|------------------------|
| Α β 42 | Α β 40 | weasured Conc. (ng/mL) |
| 0.3 | 0 | 0.3 |
| 0.3 | 0.5 | 0.8 |
| 0.3 | 2.5 | 3.0 |
| 0.3 | 10 | 6.3* |
| 0 | 0.3 | 0.3 |
| 0.5 | 0.3 | 0.8 |
| 2.5 | 0.3 | 2.8 |
| 10 | 0.3 | 6.7* |



The ability of the oligometric AB assay to measure interference from various combinations of monometric AB42 and monometric AB40 was examined. The measured ECL signal was compared for: (1) a constant level of AB42 (0.3 ng/mL) spiked with various concentrations of AB40 (0, 0.5, 2.5, and 10 ng/mL) and (2) a constant level of Aβ40 (0.3 ng/mL) spiked with various concentrations of Aβ42 (0, 0.5, 2.5, and 10 ng/mL). Varying the AB40 spike concentrations in a constant level of AB42 (0.3 ng/mL) did not impact the signal measured in the oligomeric assay. In contrast, the signal increases appreciably when increasing concentrations of AB42 are spiked into a constant level of AB40 (0.3) ng/mL). This may represent a change from monomeric AB42 to an aggregated state within the context of the experiment. The spiked levels of AB42 were >10 times the maximum physiological levels.





We present the development of two novel A β sandwich immunoassays for the measurement of either total A β or oligomeric AB. While higher order forms of AB were not detected in human CSF samples, substantial levels of total Aβ (~5–20 ng/mL) were detected in the same set of samples. The total Aβ assay was extremely sensitive and discerned a difference in total AB between normal and disease CSF. The observation that total AB levels were greater than the sum of AB40 and AB42 measured in the peptide-specific assays suggests that the total AB assay detects additional A β isoforms. We anticipate that these assays will prove useful in advancing the understanding of A β pathogenesis and in evaluating the efficacy of novel therapeutics that target A β clearance.



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| Spike Conc. (ng/mL) | | Measured Signal |
|----------------------|----------------------|-------------------|
| Α β 42 | Α β 40 | ivieasured Signal |
| 0.3 | 0 | 1048 |
| 0.3 | 0.5 | 1239 |
| 0.3 | 2.5 | 1113 |
| 0.3 | 10 | 1051 |
| 0 | 0.3 | 176 |
| 0.5 | 0.3 | 1379 |
| 2.5 | 0.3 | 5987 |
| 10 | 0.3 | 18970 |

1 Human CSF Sample Testing

Five remnant CSF pools from normal donors and 40 well-curated CSF individual samples from patients with various neurological diseases were measured using the total AB and oligomeric AB assays. Higher order AB was not detectable in the oligomeric AB assay (data not shown). In contrast, total AB was detectable in the total AB assay and assessed in terms of apparent AB40 concentration (Figure P). Total AB levels were elevated in disease CSF samples compared to normal CSF pools. The mean, median, and range for the disease CSF samples are: 10.8 ng/mL, 9.4 ng/mL, and 6.0–26.5 ng/mL, respectively. The mean, median, and range for the normal CSF pools are: 4.8 ng/mL, 4.8 ng/mL, and 3.6–6.3 ng/mL, respectively. Figure Q compares Aβ peptide concentration detected in 5 remnant CSF pools measured using several assay formats. Interestingly, the sum of the mean concentrations of AB42 and AB40 detected in the peptide-specific assays is less than the total AB measured concentration. This result suggests that the total AB assay detects additional AB-related species in human CSF.





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