

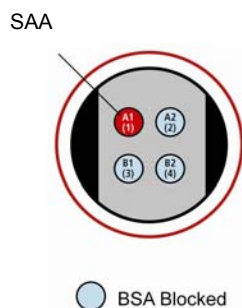
# MSD<sup>®</sup> 96-Well MULTI-ARRAY<sup>®</sup> SAA Assay

The following assay protocol has been optimized for analysis of serum amyloid A (SAA) in human serum and plasma samples.

Storage

## MSD Materials

<input type="checkbox"/> Read Buffer T (4X), with surfactant	RT
<input type="checkbox"/> Blocker A Kit	RT
<input type="checkbox"/> MULTI-SPOT <sup>®</sup> 96-well 4 Spot Human SAA Plate(s)	2-8 °C
<input type="checkbox"/> SULFO-TAG <sup>™</sup> Anti-hSAA Antibody (50X) <sup>1</sup>	2-8 °C
<input type="checkbox"/> Diluent 15	≤-10 °C
<input type="checkbox"/> Human SAA Calibrator (10 µg/mL)	≤-70 °C



The SECTOR<sup>®</sup> Imager data file will identify spots according to their well location, not by the coated capture antibody name.

<sup>1</sup> SULFO-TAG labeled detection antibodies may be light-sensitive, so they should be stored in the dark.

FOR RESEARCH USE ONLY.  
NOT FOR USE IN DIAGNOSTIC OR THERAPEUTIC PROCEDURES.



**Notes:**

## *Other Materials & Equipment (not supplied)*

- ❑ Deionized water for diluting Read Buffer
- ❑ Phosphate buffered saline (PBS) for dilution of Blocker A for sample dilution
- ❑ Phosphate buffered saline + 0.05% Tween-20 (PBS-T) for plate washing
- ❑ Adhesive plate seals
- ❑ Microtiter plate shaker
- ❑ Plate washer or other efficient multi-channel pipetting equipment for washing 96-well plates
- ❑ Liquid handling equipment for desired throughput that must accurately dispense 5, 10, 25, 40, 150, and 200  $\mu\text{L}$  into a 96-well micro plate

*Read the entire detailed instructions before beginning work.*

## *Protocol at a Glance*

**The protocol can be completed in approximately 4 hours** if each reagent is prepared during the preceding incubation. This time can be reduced to **3 hours if the blocking reagent is added the night before**. All reagents can be prepared hours ahead of time if desired.

- Step 1.** Add Blocking Solution, incubate 1 hour, wash.  
(alternatively, block plates overnight at 4 °C).
- Step 2.** Add 40  $\mu\text{L}$  of Diluent 15.  
Add 10  $\mu\text{L}$  of Samples or Calibrator, incubate 2 hours, wash.
- Step 3.** Add 25  $\mu\text{L}$  of Detection Antibody, incubate 1 hour, wash.
- Step 4.** Add 150  $\mu\text{L}$  of Read Buffer, read plate and analyze data.

## *Preparation Instructions*

### **Prepare Blocker A Kit:**

Follow instructions included with the Blocker A Kit.  
This will yield a 5% (w/v) Blocker A Solution.



## Notes:

*Diluent 15 is stable for one week at 4 °C. For longer storage, aliquot and store at ≤-10 °C. Diluent 15 may be refrozen twice.*

### Prepare Calibrator Dilutions:

Determine how many Calibrator levels and replicates will be tested. Each well will require 10 µL of Calibrator. Thaw one vial of Calibrator, and prepare the required Calibrator dilution series using Diluent 15.

- 1) A recommended Calibrator dilution procedure is listed below for 3 replicates of Calibrator concentrations spanning a wide range, plus 1 zero-Calibrator point (sample plate layout shown below).
  - Prepare a seven point calibration curve using 1/7 serial dilution as follows:
  - Dilute 10 µL of the stock Calibrator into 90 µL of Diluent 15. This yields the high Calibrator for the top of the curve (1000 ng/mL).
  - Use the high Calibrator (1000 ng/mL) to prepare serial dilutions for the rest of the curve. Add 10 µL of solution to 60 µL Diluent 15 to make a Calibrator solution at 143 ng/mL. Repeat the 1/7 serial dilution five times to make Calibrator solutions of 20, 2.9, 0.42, 0.06, and 0.008 ng/mL.
  - This will create 7 Calibrators for the analyte. The recommended 8<sup>th</sup> dilution is Diluent 15 alone (e.g. zero Calibrator).
- 2) Once the expected range of sample concentrations is known, the Calibrator concentrations can be adjusted appropriately to produce the desired standard curve.

### Prepare 1% Blocker A Solution for Sample Diluent:

1. Determine the amount of 1% Blocker A Solution needed for the experiment. Each sample requires approximately 300 µL 1% Blocker A Solution when diluting sample according to the recommendation below.
2. Dilute 5% Blocker A Solution to 1% with PBS.

### Prepare Samples:

Dilute samples 1/200 in 1% Blocker A Solution. For example a recommended two-step dilution is as follows:

- Prepare an initial 10-fold dilution by adding 10 µL of sample to 90 µL of Blocker A solution and mix thoroughly.
- Prepare the 200X diluted sample by starting with the 10X diluted sample and diluting by a factor of 20; add 10 µL of the 10X diluted sample to 190 µL of Blocker A solution.



## Notes:

### Prepare the 1X Detection Antibody Solution

- a) In a 15 mL tube combine:
- 60  $\mu$ L of 50X SULFO-TAG Anti-hSAA Antibody
  - 2.94 mL of Diluent 15
- b) This will yield 3 mL of diluted Detection Antibody Solution at the working concentration with sufficient volume for one plate.

*Detection Antibody Solution is stable at room temperature for a few hours and should be stored in the dark when not in use.*

### Dilute Read Buffer:

- In a 50 mL tube combine (per plate):
- 5 mL 4X Read Buffer T
  - 15 mL deionized water

*Diluted Read Buffer may be stored at room temperature for later use.*

## Assay Protocol

Begin with a MULTI-SPOT 96-well 4 Spot Human SAA Plate.  
No pre-treatment is necessary.

1. Add 150  $\mu$ L/well of 5% Blocker A Solution and incubate at room temperature for 1 hour with shaking (or overnight at 4 °C).
2. Wash plates 3 times with 200  $\mu$ L per well phosphate buffered saline with 0.05% Tween-20 (PBS-T).
3. Add 40  $\mu$ L/well of Diluent 15.
4. Add 10  $\mu$ L/well Calibrator or diluted sample and incubate at room temperature with shaking for 2 hours.
5. Wash plates 3 times with 200  $\mu$ L per well PBS-T.
6. Add 25  $\mu$ L/well of 1X Detection Antibody Solution and incubate at room temperature with shaking for 1 hour.
7. Wash plates 3 times with 200  $\mu$ L per well PBS-T.
8. Prepare SECTOR Imager so that plate can be read immediately after Read Buffer addition.
9. Add 150  $\mu$ L/well 1X Read Buffer T. Avoid bubbles. The use of an electronic multi-pipettor at moderate speed setting is recommended.
10. Analyze immediately with SECTOR Imager.

*Bubbles introduced to the well during Read Buffer addition will interfere with reliable imaging of the plate.*



**Sample Plate Layout:**

		1	2	3	4	5	6	7	8	9	10	11	12
ng/mL Calibrator (7-fold dilutions)	A	1000											
	B	143											
	C	20											
	D	2.9											
	E	0.42											
	F	0.06											
	G	0.008											
	H	0											
		<b>Calibrator</b>			<b>samples</b>								

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