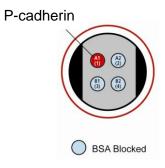
# MSD® 96-Well MULTI-ARRAY® Human P-Cadherin Assay

The following assay protocol has been optimized for analysis of P-cadherin in human serum or plasma samples.

-		Storage
MSD Materials		
	Read Buffer T (4X), with surfactant	RT
	Blocker A Kit	RT
	MULTI-SPOT® 96-well 4 Spot Human P-cadherin Plate(s)	2-8 °C
	SULFO-TAG™ Anti-hP-cadherin Antibody (50X)¹	2-8 °C
	Diluent 100	2-8 °C
	Diluent 11	≤-10 °C
	Diluent 7	≤-10 °C
	Human P-cadherin Calibrator (10 μg/mL)	≤-70 °C



The SECTOR® Imager data file will identify spots according to their well location, not by the coated capture antibody name.

FOR RESEARCH USE ONLY.
NOT FOR USE IN DIAGNOSTIC OR THERAPEUTIC PROCEDURES.



<sup>&</sup>lt;sup>1</sup> Some SULFO-TAG labeled detection antibodies may be light-sensitive, so they should be stored in the dark.

Notes:

## Other Materials & Equipment (not supplied)

- □ Deionized water for diluting Read Buffer
- □ Phosphate buffered saline for plate washing
- □ Adhesive plate seals
- □ Microtiter plate shaker
- □ Plate washer, or other efficient multi-channel pipetting equipment for washing 96-well plates
- Appropriate liquid handling equipment for desired throughput that must accurately dispense 25 and 150 μL into a 96-well microplate

Read the entire detailed instructions before beginning work.

The assay protocol was optimized for human serum samples. For significantly different sample matrices, it is recommended to use a Calibrator diluent that is similar to the sample matrix (e.g. lysis buffer + carrier protein).

#### Protocol at a Glance

The protocol can be completed in approximately 5.5 hours if each reagent is prepared during the preceding incubation. This time can reduced to 4.5 hours if the blocking reagent is added the night before.

- **Step 1.** Add Blocking solution, incubate 1 hour, wash.
- Step 2. Add 25  $\mu$ L of Diluent 7. Add 25  $\mu$ L of samples or Calibrator, incubate 2 hour, wash.
- Step 3. Add 25 µL of Detection Antibody, incubate 2 hour, wash.
- **Step 4.** Add 150 µL of Read Buffer and analyze plate.

## **Preparation Instructions**

#### Prepare Blocker A solution:

Follow instructions included with the Blocker A Kit.

#### **Prepare Calibrator dilutions:**

- Determine how many Calibrator levels and replicates will be run.
   Each well will require 25 μL of Calibrator. Thaw one vial of P-cadherin Calibrator stock solution and prepare the required Calibrator dilution series using the stock solution and Diluent 100.
  - A recommended Calibrator dilution procedure is listed below for up to 3 replicates of 7 Calibrator concentrations spanning a wide range, plus 1 zero-Calibrator point.



Notes:

- Prepare 200 μL of a high Calibrator containing 1000 ng/mL P-cadherin by adding 20 μL of the Calibrator stock solution at 10 μg/mL to 180 μL Diluent 100.
- Prepare 6 additional 1:5 serial dilutions, beginning with the high Calibrator, by adding 50 μL of the Calibrator to 200 μL Diluent 100.
- This will create 7 Calibrators with 1000, 200, 40, 8, 1.6, 0.32 and 0.064 ng/mL of P-cadherin.
- The recommended 8<sup>th</sup> dilution is Diluent 100 alone (i.e. zero Calibrator).
- ❖ Once the expected range of sample concentrations is known, the Calibrator concentrations can be adjusted appropriately.
- 2. Calibrators are stable at room temperature for at least a few hours.

#### Prepare Samples:

Dilute serum or plasma samples 10-fold in Diluent 100. Each well will require 25  $\mu$ L of diluted sample.

### Prepare Detection Antibody Reagent:

- 1. Each well requires 25  $\mu L$  of Detection Antibody Reagent. Prepare 3 mL per plate.
- 2. In a 15 mL tube combine:
  - a. 2.94 mL Diluent 11
  - b. 60 μL of 50X SULFO-TAG Anti-hP-cadherin Antibody (final concentration: 1X)

#### Prepare Read Buffer Solution:

In a 50 mL tube combine (per plate):

- □ 5 mL 4X Read Buffer T
- □ 15 mL deionized water

Diluted Read Buffer may be stored at room temperature for later use.



## **Assay Protocol**

Begin with a MULTI-SPOT 96-well 4 Spot Human P-cadherin plate. No pre-treatment is necessary.

- 1. Add 150 μL/well of Blocker A and incubate at room temperature for 1-2 hours.
- 2. Wash plates 3 times with phosphate buffered saline (PBS)
- 3. Add 25  $\mu$ L/well of Diluent 7. Add 25  $\mu$ L/well Calibrator or 10-fold diluted sample and incubate at room temperature with shaking for 2 hours.
- 4. Wash plates 3 times with PBS.
- 5. Add 25  $\mu$ L/well Detection Antibody Reagent and incubate at room temperature with shaking for 2 hours.
- 6. Wash plates 3 times with PBS.
- 7. Prepare SECTOR Imager so that plate can be read immediately after Read Buffer addition.
- 8. Add 150 μL/well 1X Read Buffer T. Avoid bubbles.

Bubbles introduced to the well during Read Buffer addition will interfere with reliable imaging of the plate.

9. Analyze immediately with SECTOR Imager.

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