### Materials List

<table>
<thead>
<tr>
<th>Material</th>
<th>Vendor</th>
<th>Product</th>
<th>Catalog Number</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blocker A</td>
<td>Proliant</td>
<td>Bovine Serum Albumin, Reagent Grade Pure Powder</td>
<td>68700</td>
</tr>
<tr>
<td>Blocker B</td>
<td>Amersham Biosciences</td>
<td>Membrane Blocking Agent</td>
<td>RPN2125</td>
</tr>
<tr>
<td>Blocker C</td>
<td>BioRad</td>
<td>1X TBS/Casein Blocker</td>
<td>161-0782</td>
</tr>
<tr>
<td>Blocker D-M</td>
<td>Rockland</td>
<td>Mouse Gamma Globulin</td>
<td>D609-0100</td>
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<tr>
<td>Blocker D-R</td>
<td>Rockland</td>
<td>Rabbit Gamma Globulin</td>
<td>D610-1000</td>
</tr>
<tr>
<td>Blocker D-G</td>
<td>Equitech-Bio, Inc.</td>
<td>Goat IgG</td>
<td>SLG66-0010</td>
</tr>
<tr>
<td>Blocker D-B</td>
<td>Equitech-Bio, Inc.</td>
<td>Bovine IgG</td>
<td>SLB66</td>
</tr>
<tr>
<td>Phosphatase Inhibitor I</td>
<td>Sigma-Aldrich, Inc.</td>
<td>Phosphatase Inhibitor Cocktail 3</td>
<td>P0044</td>
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<tr>
<td>Phosphatase Inhibitor II</td>
<td>Sigma-Aldrich, Inc.</td>
<td>Phosphatase Inhibitor II</td>
<td>P-5726</td>
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<tr>
<td>Protease Inhibitor Solution</td>
<td>Pierce Biotechnology</td>
<td>Product Halt™ Protease Inhibitor Cocktail, EDTA-Free</td>
<td>78439 (10 mL, sufficient to treat 1.0 L of sample)</td>
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<td>78437 (5 mL, sufficient to treat 500 mL of sample)</td>
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<td>87785 (1 mL, sufficient to treat 100 mL of sample)</td>
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<tr>
<td>Spin Columns</td>
<td>Thermo Scientific</td>
<td>ZEBA™ Spin Desalting Column (5 columns/pack)</td>
<td>87728</td>
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<td>ZEBA Spin Desalting Column (25 columns/pack)</td>
<td>87769</td>
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### Buffer Preparation Instructions

<table>
<thead>
<tr>
<th>Buffer Type</th>
<th>Composition</th>
<th>Store Temperature</th>
<th>Notes</th>
</tr>
</thead>
</table>
| **Tris Wash Buffer (10X)** | 500 mM Tris, pH 7.5  
1.5 M NaCl  
0.2% Tween-20 | 2-8ºC. |  |
| **Tris Lysis Buffer (1X, Incomplete)** | 150 mM NaCl  
20 mM Tris, pH 7.5  
1 mM EDTA  
1 mM EGTA  
1% Triton-X-100 | 2-8ºC. |  |
Blocker A

**General Notes:**
Blocker A is added to Tris Wash Buffer (1X) to prepare Blocking Solution A and Antibody Dilution Buffer. Please see individual assay inserts for specific preparation instructions.

Blocker B

**General Notes:**
Blocker B solution is required for the dilution of cell lysates only for certain targets. Please see individual assay inserts for specific preparation instructions.

Blocker D-M

**General Notes:**
Store Blocker D-M for up to 2.5 years at ≤ -10°C. The MSD working assay concentration is 0.1%.

**Reagents Required:**
Sterile H₂O + 0.05% Sodium Azide

**Protocol:**
Reconstitute Mouse Gamma Globulin (Rockland D609-0100) by adding 5 mL H₂O + 0.05% Sodium Azide to one 0.1 g vial to prepare a 2% stock solution. Aliquot and store at ≤ -10°C or 2-8°C.

Blocker D-R

**General Notes:**
Store Blocker D-R for up to 2.5 years at ≤ -10°C. The MSD working assay concentration is 0.1%.

**Reagents Required:**
Sterile H₂O + 0.05% Sodium Azide

**Protocol:**
Reconstitute Rabbit Gamma Globulin (Rockland D610-1000) by adding 10 mL H₂O + 0.05% Sodium Azide to one 1.0 g vial to prepare a 10% stock solution. Centrifuge 3 times for 30 minutes at 2000 rpm with the supernatant removed each time and pellet discarded. Aliquot and store at ≤ -10°C or 2-8°C.
| Blocker D-B | General Notes:  
Store Blocker D-B for up to 2.5 years at ≤ -10°C.  
The MSD working assay concentration is 0.7% for MSD Mouse Cytokine Assays, and 0.1% for MSD Alzheimer's Disease Assays.  
Reagents Required:  
Sterile H₂O + 0.05% Sodium Azide  
Protocol:  
Reconstitute Bovine IgG (Equitech-Bio SLB66) by adding 1 mL H₂O + 0.05% Sodium Azide to one 0.1 g vial to prepare a 10% stock solution. Aliquot and store at ≤ -10°C or 2-8°C. |
| Blocker D-G | General Notes:  
Store Blocker D-G for up to 2.5 years at ≤ -10°C.  
The MSD working assay concentration is 0.1%.  
Reagents Required:  
Sterile H₂O + 0.05% Sodium Azide  
Protocol:  
Reconstitute Goat IgG (Equitech-Bio SLG66-0010) by adding 20 mL H₂O + 0.05% Sodium Azide to 2 g to prepare a 10% stock solution. Aliquot and store at ≤ -10°C or 2-8°C. |
**Whole Cell Lysate Preparation Instructions**

**General notes:**
All manipulations should be performed on ice. Prepare desired amount of Complete Lysis Buffer immediately prior to use. Lysis volumes will vary between cell types. Larger cells (such as NIH3T3, HeLa) should be lysed at concentrations of 1-5 x 10^6 cells per mL of Lysis Buffer. Smaller cells (such as Jurkat) should be lysed at concentrations of 1-5 x 10^7 cells per mL of Lysis Buffer.

**Reagents required:**

| Tris Lysis Buffer (1X, Incomplete) | 150 mM NaCl  
20 mM Tris, pH 7.5  
1 mM EDTA  
1 mM EGTA  
1% Triton-X-100 |
|------------------------------------|---|
| Complete Lysis Buffer | 10 mL 1X Tris Lysis Buffer  
100 µL Phosphatase Inhibitor I  
100 µL Phosphatase Inhibitor II  
100 µL Protease Inhibitor Solution |

**NOTE:** For assays that require PMSF, a final concentration of 2 mM in Complete Lysis Buffer is recommended. For 10 mL Complete Lysis Buffer, add 40 µL PMSF (from 500 mM stock prepared in DMSO) (Sigma, Catalog # P-7626). When adding PMSF to the buffer, all reagents should be at room temperature prior to mixing. The Complete Lysis Buffer should be mixed at room temperature on a rotator for 5 minutes (with no obvious precipitates), and should then be thoroughly chilled prior to use for lysate dilution or preparation.

**When analyzing cells for p38 alone, the following Lysis Buffer affords optimal performance:**

| Diluent 20 | 50 mM NaCl  
20 mM Tris, pH 7.5  
1 mM EDTA  
1 mM EGTA  
1% Triton X-100 |
|------------|---|

Prepare Complete Diluent 20 as shown above, substituting the 10 mL of Tris Lysis Buffer (1X) with Diluent 20.
Whole Cell Lysate Preparation Instructions (continued)

**Protocol:**
Cells should be prepared as desired to activate target protein.

| Suspension cells | Pellet cells (500 x g, 3 minutes at 2-8°C) and wash one time with cold 1X PBS. Pellet cells again and resuspend in 1X Complete Lysis Buffer at 1-5 x 10⁷ cells per mL. Incubate on ice for 30 minutes (a shorter incubation time of 15 minutes may be adequate for many targets). Centrifuge lysates at greater than or equal to 10,000 x g, 2-8°C for 10 minutes to clear cellular debris from the lysate. Lysates can be quantitated using a detergent compatible protein assay such as BCA. Unused lysates should be aliquoted and snap frozen and stored at -80°C. |
| Adherent cells | All volumes are determined for cells plated in 15 cm dishes. Remove media from the plates and wash cells one time with 5 mL cold 1 X PBS. Add 2 mL 1 X PBS to the plates and scrape the cells from the surface of the dish and transfer into 15 mL conical tubes. Pellet the cells at 500 x g for 3 minutes at 2-8°C. Resuspend the cells in 0.5-2 mL of Complete Lysis Buffer per dish. Alternatively after medium removal, cells can be washed one time with PBS including a careful aspiration of residual PBS and lysed directly on the dish by adding 1-2 mL (depending on cell type) of Complete Lysis Buffer per dish. Incubate on ice for 30 minutes (a shorter incubation time of 15 minutes may be adequate for many targets). Centrifuge lysates at greater than or equal to 10,000 x g, 2-8°C for 10 minutes to clear cellular debris from the lysate. Lysates can be quantitated using a detergent compatible protein assay such as BCA. Unused lysates should be aliquoted and snap frozen and stored at -80°C. |

| 96 well format modifications | Successful adaptation to a 96 well format is cell type and target-dependent. The number of cells to be plated per well should be determined per cell type. General recommended plating concentrations for adherent cells range from 1 x 10⁴-5 x 10⁴ cells per well, and approximately 2 x 10⁶ cells per mL (50-75 µL per well) for suspension cells. These numbers are provided as a guide and the optimal concentrations will vary depending upon cell line used. |
| Suspension cells | For flat bottom plates, experiments should be designed such that the final volume per well is 50-75 µL. Perform cell lysis using a 4X Complete Lysis Buffer concentrate, supplemented with protease and phosphatase inhibitors at 4X concentrations. Add 4X Complete Lysis Buffer directly to cells in the growth medium for a final 1X concentration in the well. NOTE: With some effort, a 10X Complete Lysis Buffer can also be prepared. (For conical microwell plates, perform lysis by pelleting the cells, removing most of the growth medium and adding a constant amount of 1X Complete Lysis Buffer.) |
| Adherent cells | Plate cells on biologically treated tissue culture ware to reduce variability due to cells lost as growth medium is removed. Treat cells as desired. Gently aspirate growth medium from microwell plate. A PBS wash step is not required and can introduce variability. Add 50-100 µL 1X Complete Lysis Buffer per well. Cell lysis time should be determined by the end user. Some targets are immediately available for detection, while others may require an incubation step at room temperature, 2-8°C, or on ice with gentle agitation. Carefully pipet 25-90 µL cell lysate onto prepared capture plate. It is important to transfer a constant volume and avoid pipeting too vigorously, as the introduction of air bubbles may result. |

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